

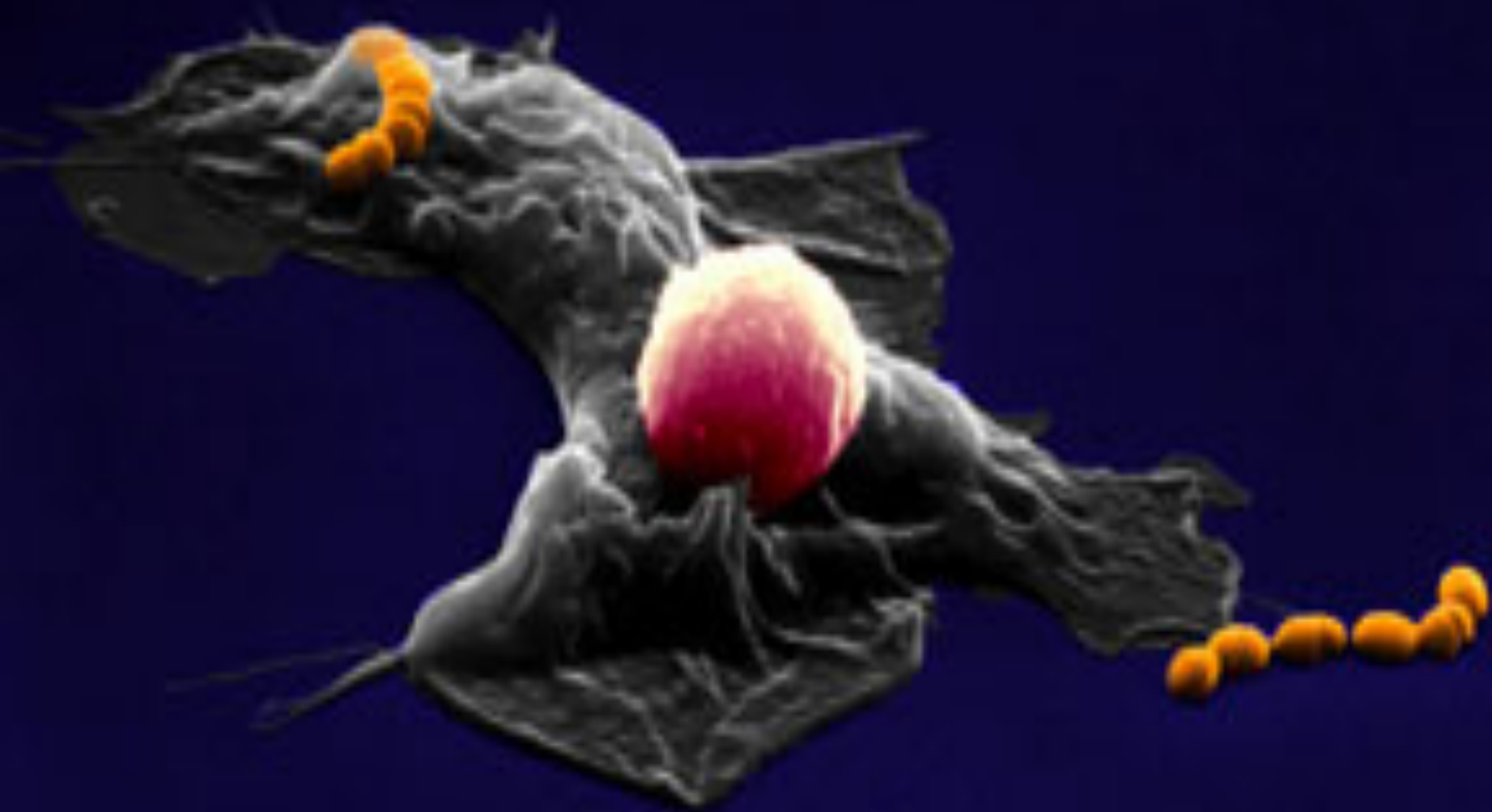
# How to monitor and mitigate immunogenicity during early phase clinical trials

**GEOFF HALE**



# Summary

- T cells orchestrate the immune response
- The established guidelines for measurement of immunogenicity are insane
- A surprisingly high proportion of healthy people have antibodies against PEG
- It may be possible to tolerise patients to prevent the formation of anti-drug antibodies



# Antigen Processing



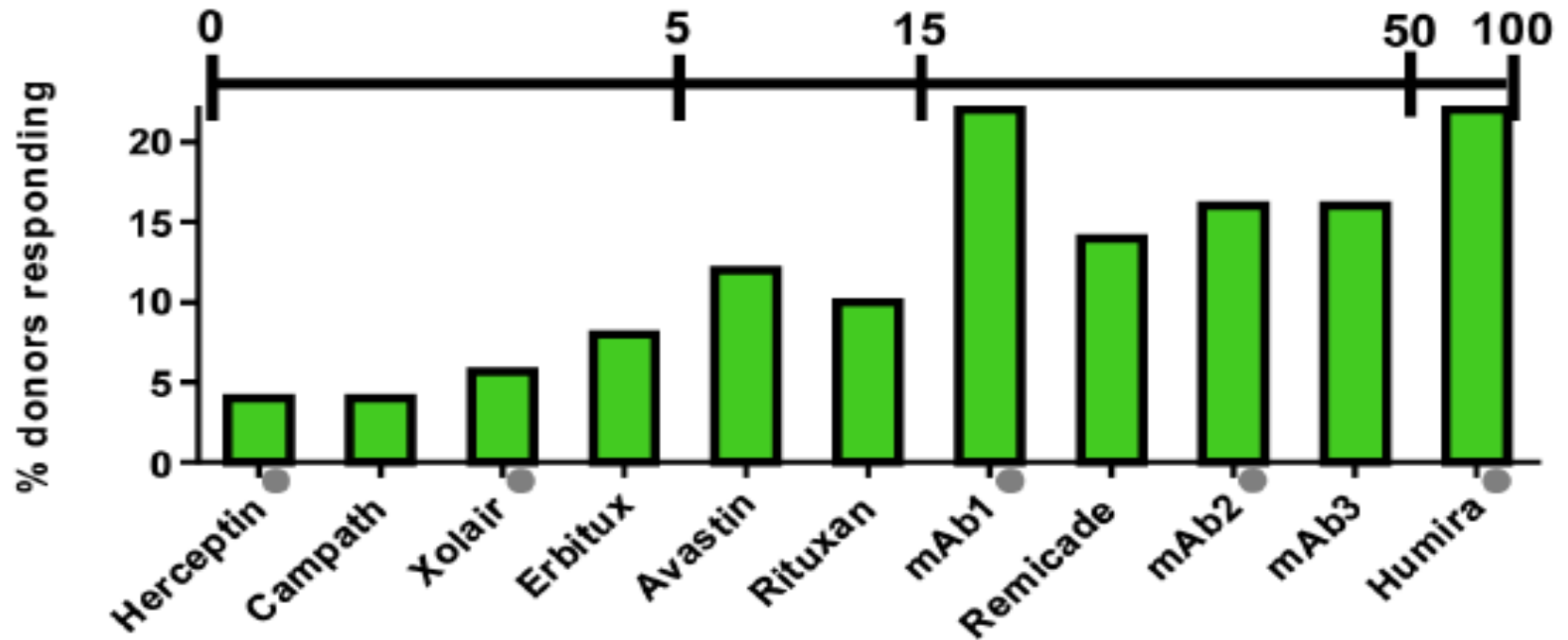
RESEARCH ARTICLE

# Use of *In Vitro* Assays to Assess Immunogenicity Risk of Antibody-Based Biotherapeutics

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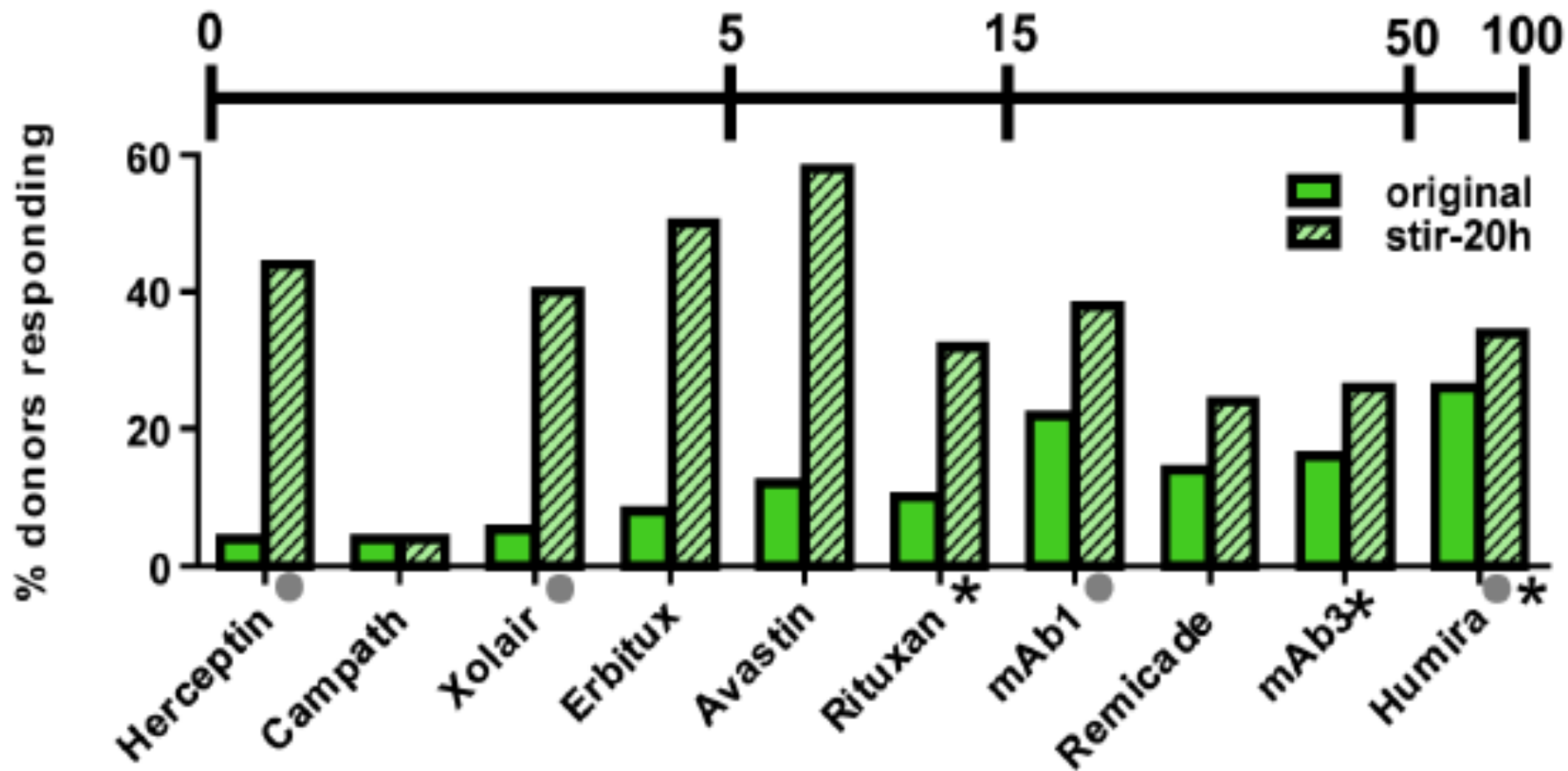
# Frequency of responses correlates with frequency of anti-drug responses in the clinic

**C** Proliferation or IL-2 Concentration



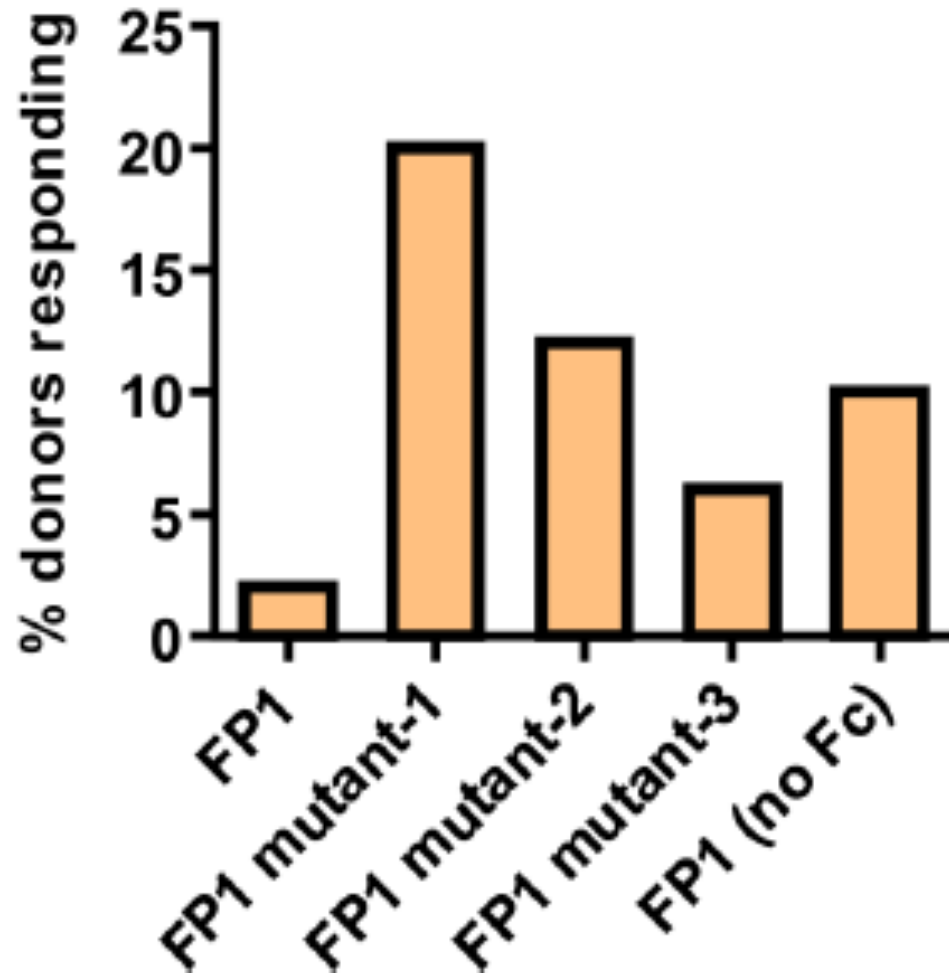
# Frequency of responses increases with aggregation

**C** Proliferation or IL-2 Concentration



# Response can be sensitive to single amino acid change

## C Proliferation + No. IL-2 Secreting Cells





## Industry White Papers

**Mire-Sluis AR et al.** Recommendations for the design and optimization of immunoassays used in the detection of host antibodies against biotechnology products.

*J. Immunol. Meth.* 289:1-16 (2004)

**Shankar et al.** Recommendations for the validation of immunoassays used for the detection of host antibodies against biotechnology products.

*J. Pharm Biomed Anal.* 48:1267-1281 (2008)

## Regulatory Guidance

**EMA:** Guideline on immunogenicity assessment of therapeutic proteins.

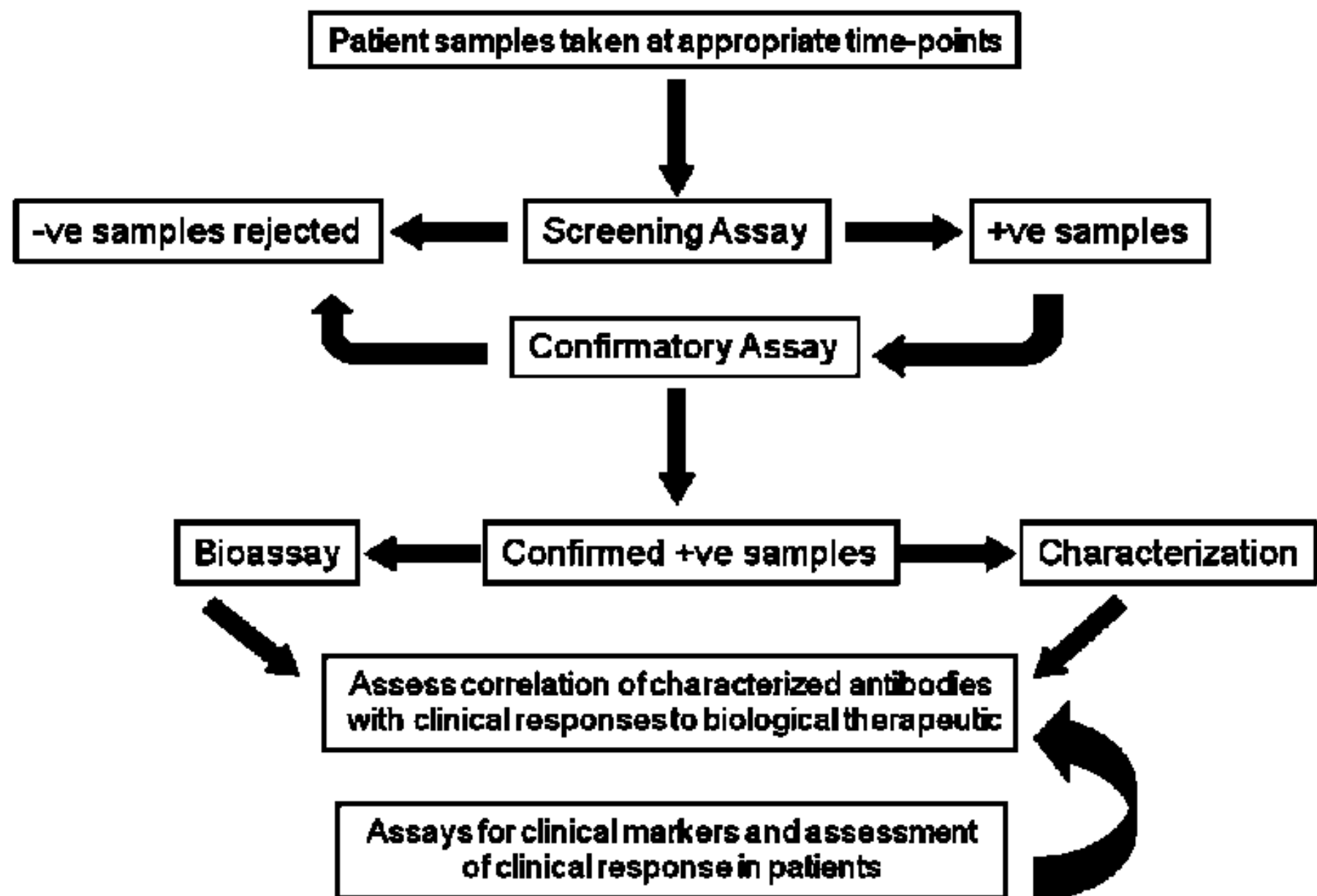
EMA/CHMP/BMWP/14327/2006

**EMA:** Guideline on immunogenicity assessment of monoclonal antibodies

EMA/CHMP/BMWP/86289/2010

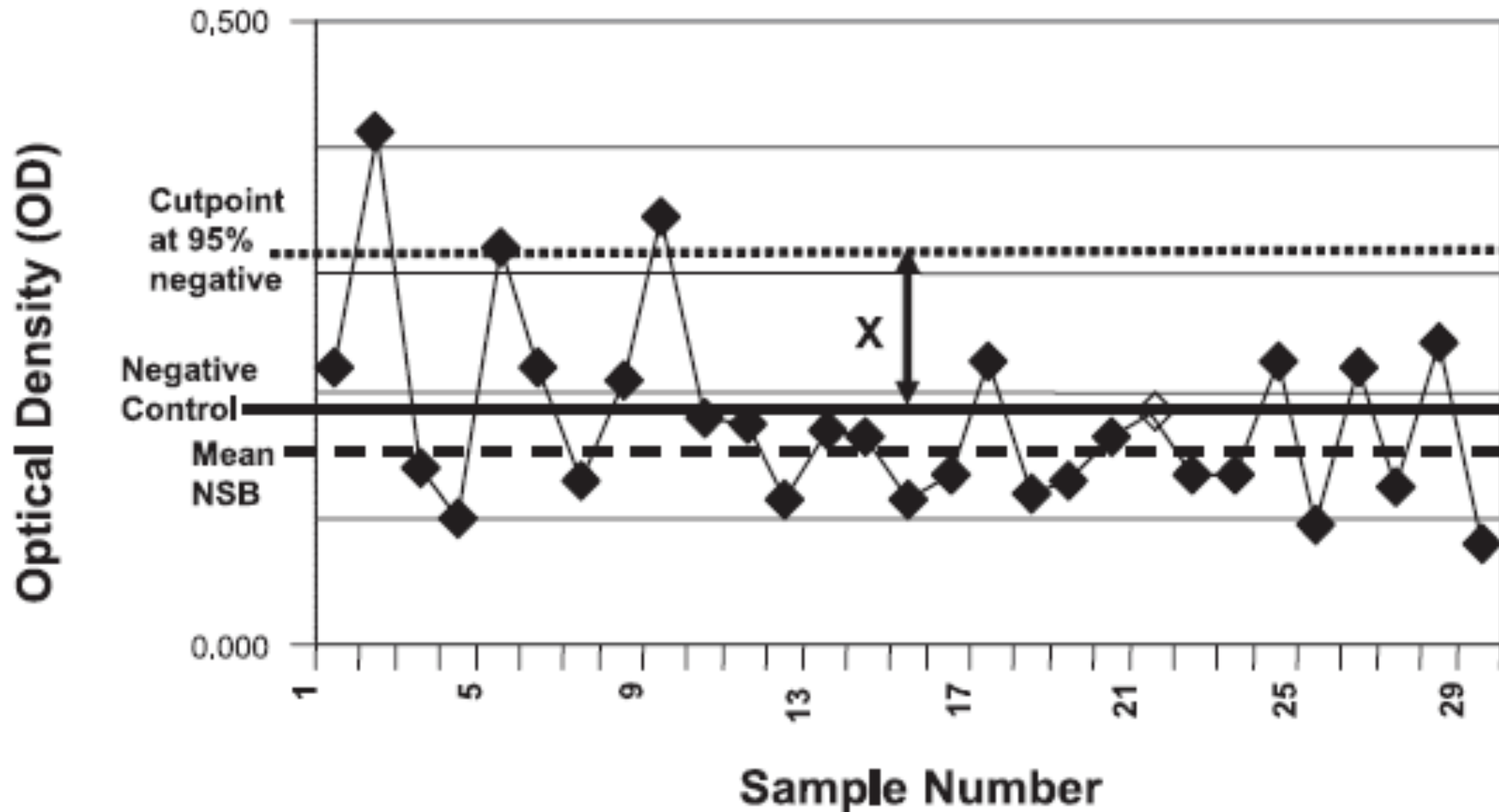
**FDA:** Guidance for Industry: Immunogenicity Testing of Therapeutic Proteins (2014)

**FDA:** Guidance for Industry: Assay Development and Validation for Immunogenicity Testing of Therapeutic Protein Products (draft, 2016)



# Determination of cut-point

*A.R. Mire-Sluis et al. / Journal of Immunological Methods 289 (2004) 1–16*



# Screening Assay: Cut point

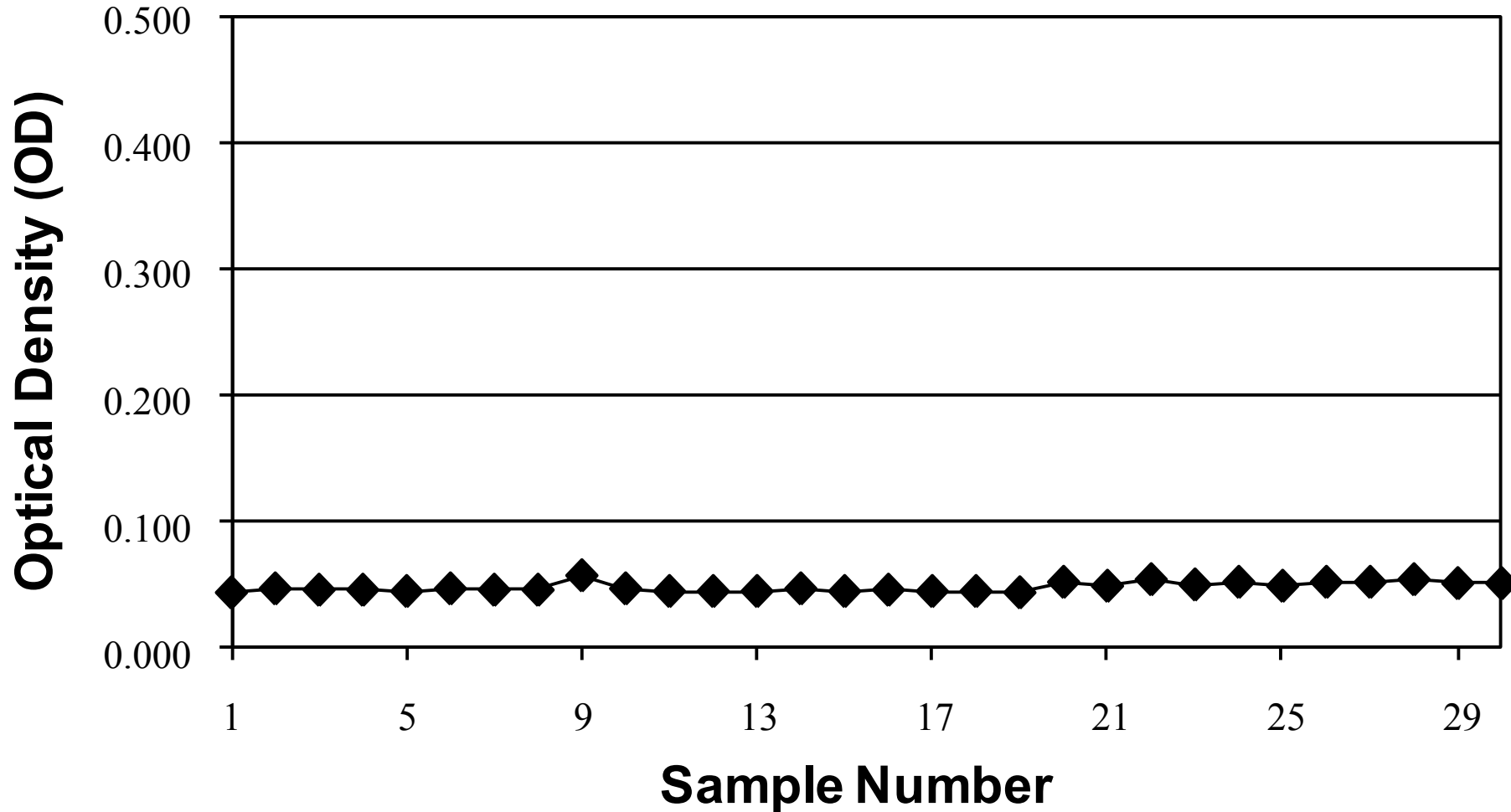
AAPS	EMA	FDA
It is appropriate to have 5% false positives	Detection of some false positive results is inevitable	recommends ... a 5% false positive rate.  The approach ... will depend on various factors

# Screening Assay: Sensitivity

AAPS	EMA	FDA
Strive for sensitivities near 250 to 500 ng/mL	Capable of detecting antibodies in all antibody-positive samples/patients	Traditionally recommended at least 250 to 500 ng/mL Now recommends at least 100 ng/mL

# Determination of cut-point

*Data from Ingrid Caras, PDL*



# Case Study 1: What Happened

- ◆ Ran post-dose Study samples
- ◆ Applied cut point (OD ~0.06)
- ◆ High incidence of 'positives' with low OD values
  - a few hundredths of OD unit above cut point
- ◆ Most were sporadic or 'transient'
  - Pos at one time point and Neg at the next
- ◆ Not typical of a real immune response
- ◆ Impossible to confirm in a competition assay
  - OD's too close to the floor

## *Subject Example*

Day	OD	
0	0.067	
14	0.087	<i>Pos</i>
28	0.074	
56	0.076	<i>Pos</i>
84	0.066	



# Case Study 1: Conclusion

- ◆ Strict adherence to the statistical approach rules would mean reporting these subjects as *positive* – resulting in high incidence
- ◆ BUT
  - *Most positives based on few hundredths of OD unit above cut point*
  - *Sporadic timing didn't look like a real immune response*
  - *Previous studies showed very low immunogenicity*
- ◆ Important to apply some common sense
- ◆ Combination of tight assay with v low background and v low SD resulted in cut point that was too low
- ◆ Result – we pick up a lot of 'noise'

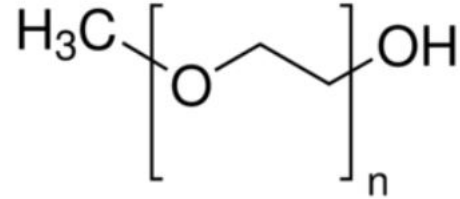
## *Subject Example*

Day	OD	
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# Cut Point and Sensitivity

Insane	Sane
5% false positive	Minimise false positive and false negative.
Arbitrary sensitivity based on assay variability	Optimal sensitivity (100 ng/mL) based on likelihood of clinical sequelae
Impossible to compare different assays	Assay results can be compared
Assay development and validation is exceedingly cumbersome	Assay development and validation is simple
Any sort of in house reference might be used	Reference should be optimised and well characterised

# Polyethylene glycol



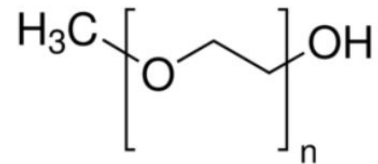
(aka polyethylene oxide, polyoxyethylene)

**Pharmaceutical:** laxative, eye drops, excipient, protein modifier

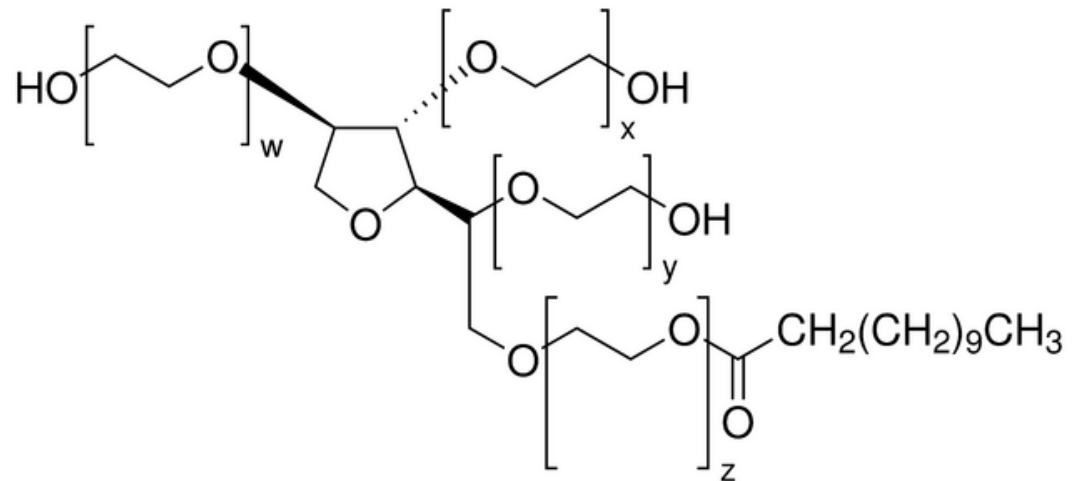
**Commercial:** wood preservatives, paints, rocket fuel, gas scrubber in power plants, anti-foaming agent, ceramic manufacture

**Domestic:** tooth paste, skin creams, lubricants, inkjet printers, paintballs, anti-foaming agent in food, e-cigarettes

# Detergents containing polyethoxy groups



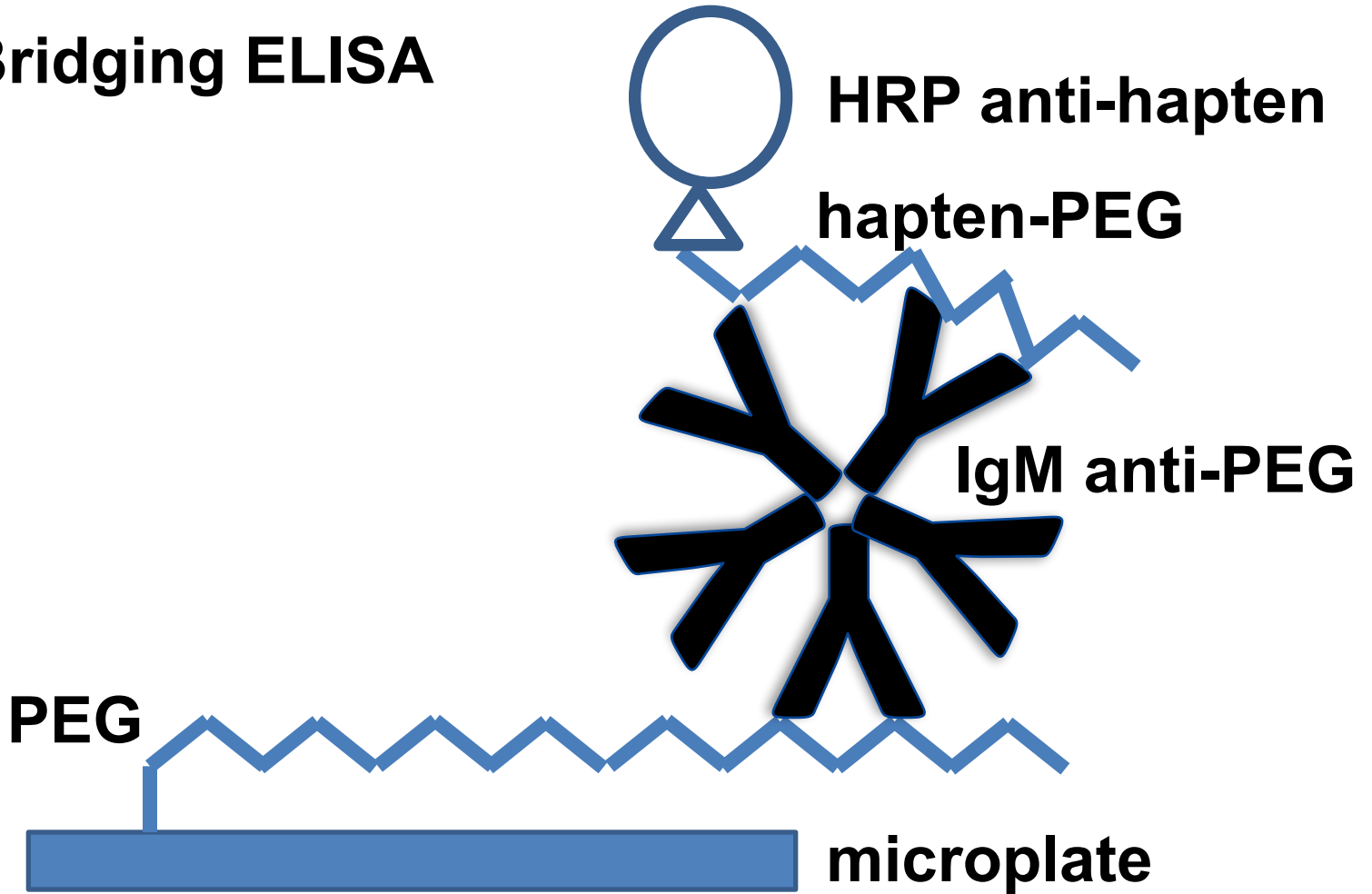
Tween 20



also:

Nonidet, Pluronic, Polysorbate, Triton

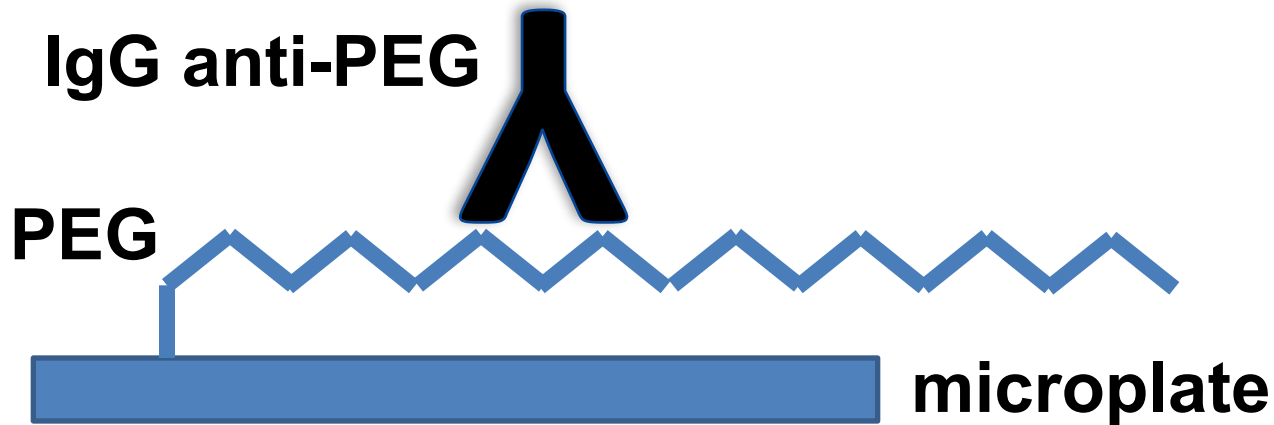
# Bridging ELISA



# Bridging ELISA



**Detection not possible!**



# Direct ELISA

HRP anti-IgG

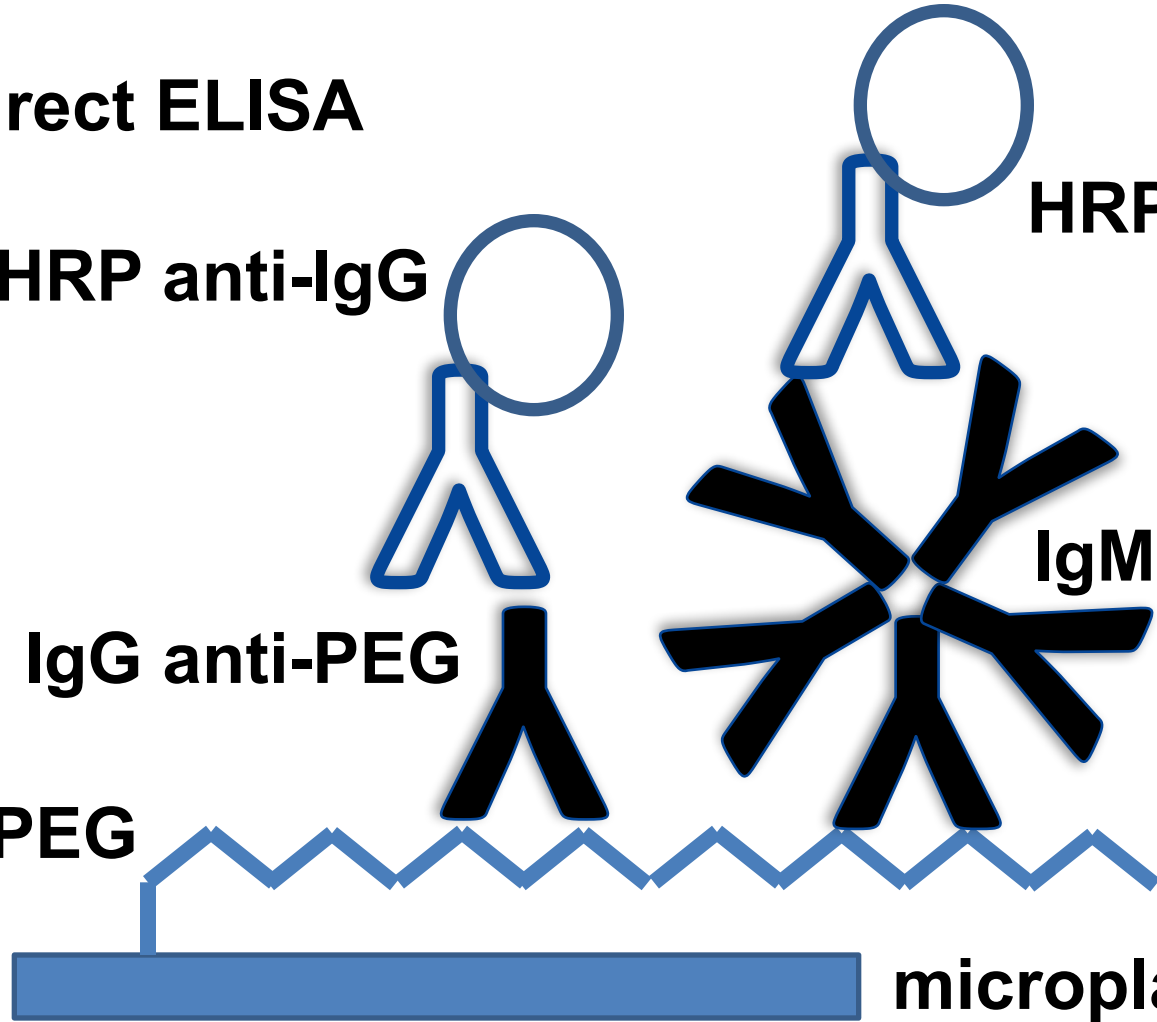
HRP anti-IgM

IgG anti-PEG

IgM anti-PEG

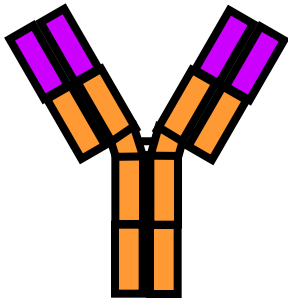
PEG

microplate

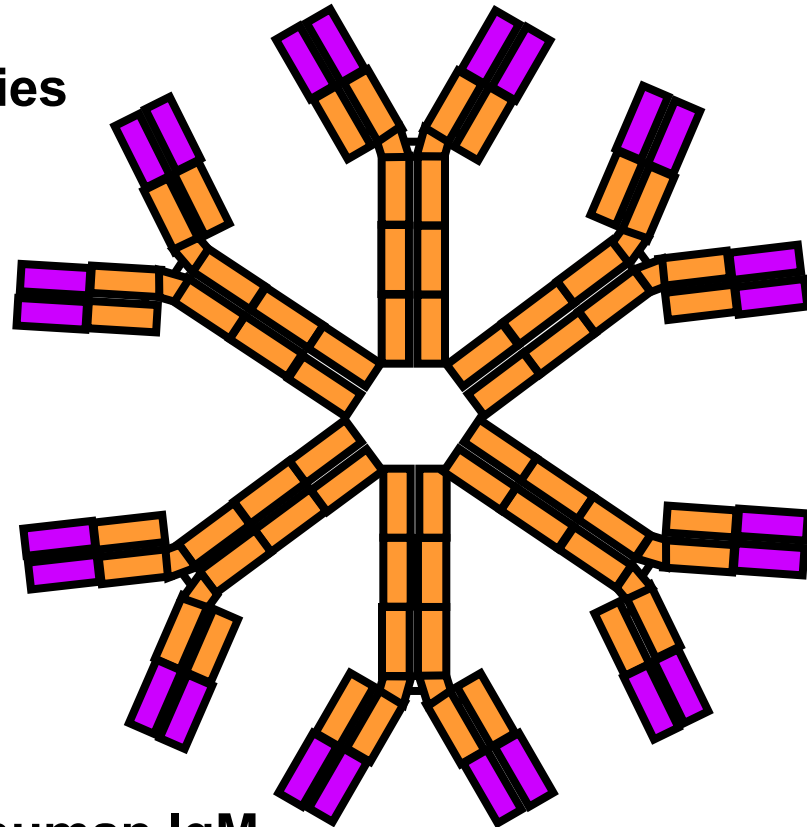




**Anti-PEG control antibodies**



**chimeric human IgG1**



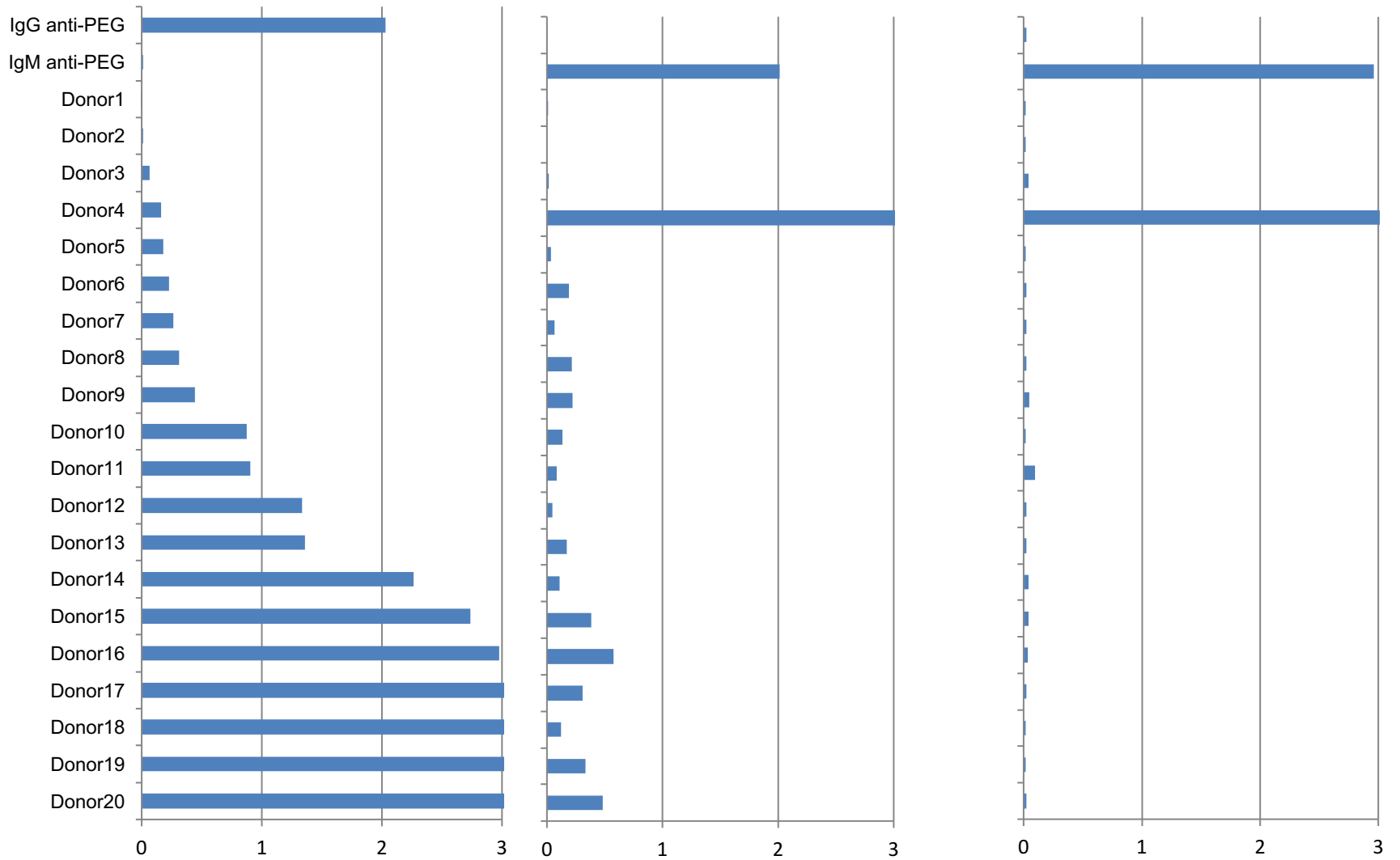
**chimeric human IgM**



### IgG-direct ELISA

### IgM-direct ELISA

### Bridging ELISA



## Proportion and titre of anti-PEG antibodies in 50 healthy individuals

Titre	Number (%)	
	IgG	IgM
Negative	9 (18%)	20 (40%)
Positive	41 (82%)	30 (60%)
10-20	21	20
40	0	2
80	0	2
160	3	2
320	3	0
640	6	1
1280 or greater	8	3

## Effect of Tween in wash buffer

	Absorbance			
Sample	IgG anti-PEG		IgM anti-PEG	
	PBS	PBS-Tween	PBS	PBS-Tween
<b>M-A05</b>	3.187	0.020	2.886	3.149
<b>M-A07</b>	3.015	-0.049	0.151	0.379
<b>M-A09</b>	2.967	0.004	0.439	0.598
<b>F-A03</b>	3.095	0.062	-0.194	0.175
<b>F-A05</b>	3.008	-0.035	-0.013	0.076
<b>IgG control</b>	1.142	0.502		
<b>IgM control</b>			1.484	1.987

# Conclusions

- Unexpectedly high frequency and titre of IgG anti-PEG antibodies in healthy donors
- Previous assays would not have detected them
- Consequence of exposure to PEG in the environment?
- Implications for PEG in drug conjugates?
- Utility of recombinant control reagents
- Beware of using bridging assay if the antigen has repeating epitopes
- Traditional cut-point approach cannot be used when there are pre-existing antibodies

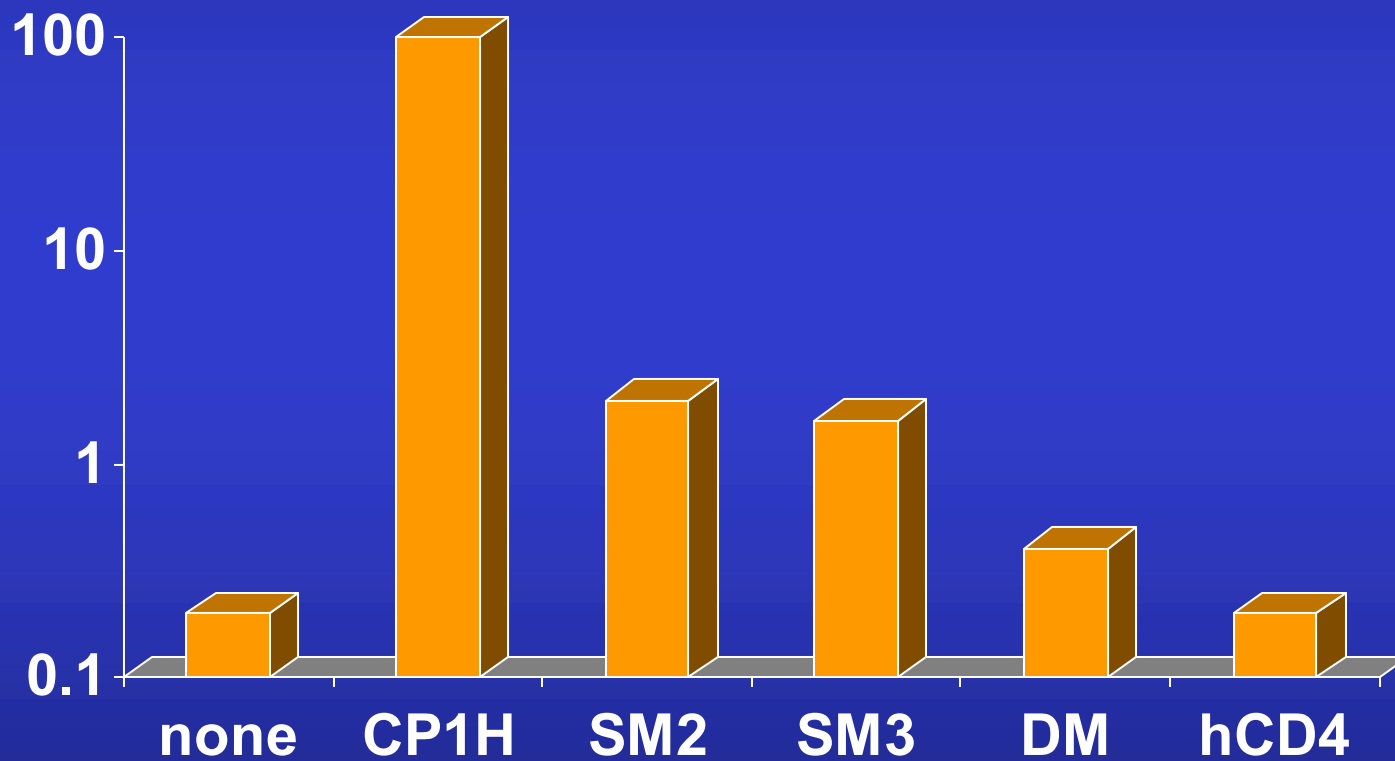
# Prevention of unwanted antibodies by induction of tolerance

- High-zone tolerance to deaggregated IgG
- High response to cell-binding antibodies
- Tolerance induced by non-binding antibodies

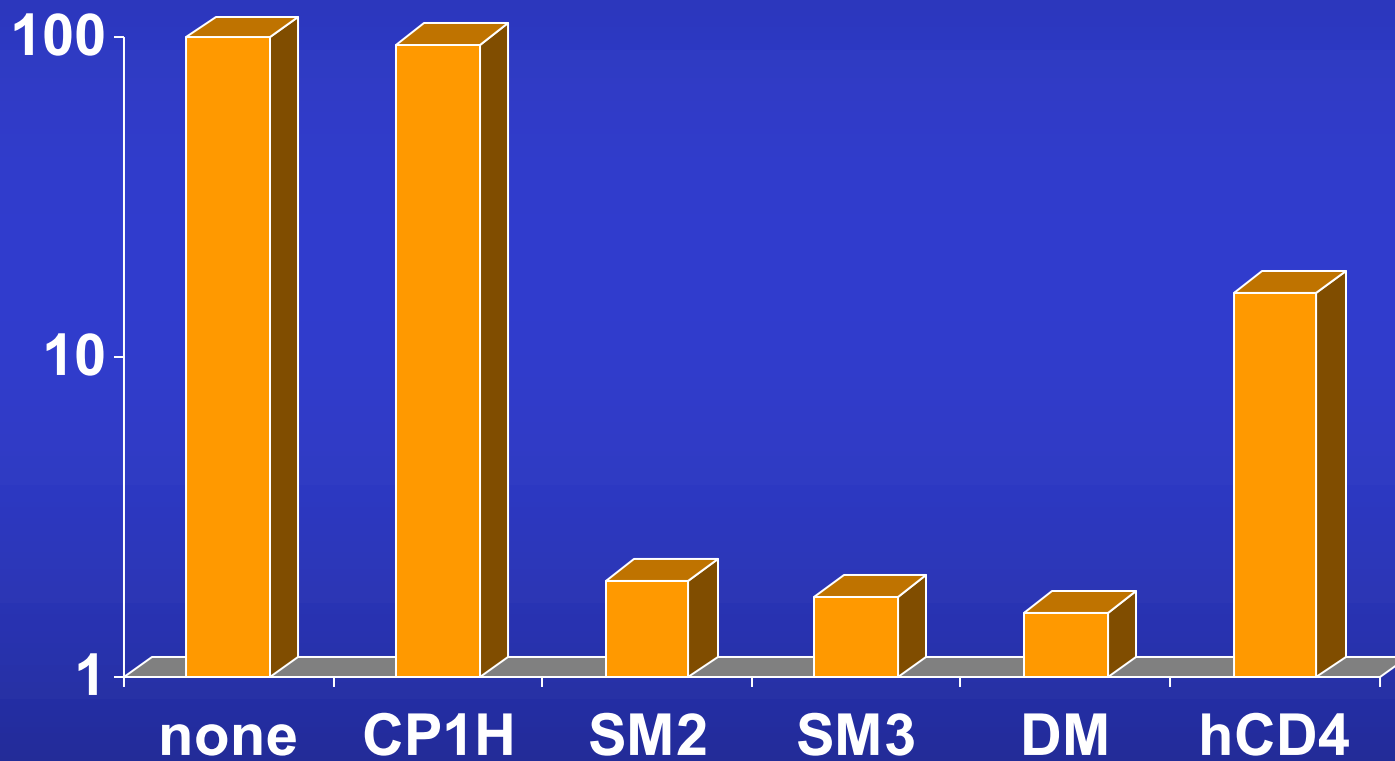
## alemtuzumab mutants

	V <sub>H</sub> CDR2 sequence	Binding
<b>Wild-type</b>	<b>R D K A K G</b>	<b>100 %</b>
<b>SM1</b>	<b>* * D * * *</b>	<b>50%</b>
<b>SM2</b>	<b>* K * * * *</b>	<b>10%</b>
<b>SM3</b>	<b>* * * * D *</b>	<b>&lt; 1%</b>
<b>DM</b>	<b>* * D * D *</b>	<b>&lt; 1%</b>

# Immunogenicity of mutants in vivo



# Responses to wild-type alemtuzumab





# Clinical Trial of SM3 to induce tolerance

- Proof of concept study in 15 patients
- High dose SM3 followed by standard course of treatment with alemtuzumab (5 days)
- Second cycle of treatment after 12 months

